

FIDIS™ EXTRACELLULAR PROTEIN BUFFER REAGENT KIT

English

REF

MX HB 0001



RUO

INTRODUCTION

Advances in the field of cell biology have defined a complex and interdependent set of extracellular and intracellular signaling molecules that control normal cell function. Therefore, there is growing interest among clinicians as well as drug discovery groups in simultaneous monitoring multiple components of signaling pathways. Solid phase multiplex protein assays have become the primary tools of choice in these studies as they maximize efficiency by permitting several analytes to be profiled within individual samples.

bmd's Multiplex Bead Immunoassays are solid phase protein assays which use spectrally encoded antibody-conjugated beads as the solid support. The spectral beads are suitable for use in singleplex assays or may be mixed for multiplex assays according to the researcher's requirements. Each assay is carefully designed and tested to assure that sensitivity, range and correlation are maximized. The assay is performed in a 96 well plate format and analyzed with a FIDIS™ instrument, which monitors the spectral properties of the capture beads while simultaneously measuring the quantity of associated fluorophore. Standard curves generated with this assay system extend over several orders of magnitudes of concentrations, while the sensitivity and quantitation of the assays are comparable to ELISAs (Enzyme Linked-Immuno-Sorbent Assays). Assay standards are calibrated to NIBSC reference preparations, when available, to assure accurate and reliable results.

FIDIS™ Human Extracellular Antibody Bead Kit and **FIDIS™ Antibody Bead Kits** are intended for the *in vitro* quantitative determination of analytes in serum, plasma, and tissue culture medium.

This kit has been configured for research use only and is not to be used in diagnostic procedures.

Read entire protocol before use.

PRINCIPLE OF THE METHOD

bmd's Multiplex Bead Immunoassays are solid phase sandwich immunoassays, which are designed to be analyzed with a FIDIS™ instrument. The spectral properties of 100 distinct bead regions can be monitored with the FIDIS™ instrument, a capability that affords this assay system the potential for measuring up to 100 different analytes in a single sample.

- Beads of defined spectral properties conjugated to analyte specific capture antibodies and samples (including standards of known analyte concentration, control specimens, and unknowns) are pipetted into the wells of a filter bottom microplate and incubated for 2 hours. During this first incubation, analytes bind to the capture antibodies on the beads.
- After washing the beads, analyte-specific biotinylated detector antibodies are added and incubated with the beads for 1 hour. During this second incubation, the analyte-specific biotinylated detector antibodies recognize their epitopes and bind to the appropriate immobilized analytes.
- After removal of excess biotinylated detector antibodies, streptavidin conjugated to the fluorescent protein, R-Phycoerythrin (Streptavidin-RPE), is added and incubated for 30 minutes. During this final incubation, the Streptavidin-RPE binds to the biotinylated detector antibodies associated with the immune complexes on the beads, forming a four-member solid phase sandwich.
- After washing to remove unbound Streptavidin-RPE, the beads are analyzed with the FIDIS™ instrument. By monitoring the spectral properties of the beads and the amount of associated R-Phycoerythrin (RPE) fluorescence, the concentration of one or more analytes can be determined.

MATERIAL REQUIRED BUT NOT PROVIDED

- FIDIS™ system. Please contact [bmd](#) for instrument and software placement services.
- [bmd](#) filtration manifold for bead washing.
- [bmd](#) data analysis and graphing software program.
- Sonicating water bath (e.g., Cole Parmer, Cat. # 08849-00).
- Orbital shaker (e.g., Fisher, Cat. # 14-271-9).
- Calibrated, adjustable, precision pipettes, preferably with disposable plastic tips. (A manifold multi-channel pipette is desirable.)
- Distilled or deionized water.
- Glass or plastic tubes.
- Beakers and graduated cylinders in various sizes.
- Aluminum foil.
- Extra filter plate covers.

REAGENTS PROVIDED

| | MX HB 0001 |
|--|------------|
| <i>Filter Plate, 96 wells per plate</i> | 1 plate |
| <i>Wash Solution Concentrate (20x)</i> Contains 0.1% sodium azide; (15 mL per bottle) | 1 bottle |
| <i>Assay Diluent.</i> Contains 0.1% sodium azide; (15 mL per bottle) | 1 bottle |
| <i>Incubation Buffer.</i> Contains 0.1% sodium azide; (12 mL per bottle) | 1 bottle |
| <i>Biotin Diluent.</i> Contains 3.3 mM thymol; (12 mL per bottle) | 1 bottle |
| <i>Streptavidin-RPE Concentrate (10x).</i> Contains 0.1% sodium azide; (1 mL per vial) | 1 vial |
| <i>Streptavidin-RPE Diluent.</i> Contains 3.3 mM thymol; (12 mL per bottle) | 1 bottle |

STABILITY AND STORAGE

- Store reagents and assay strips at +2°C/+8°C in their own package.
- Do not freeze

SAFETY

All blood components and biological materials should be handled as potentially hazardous. Follow universal precautions as established by the Centers for Disease Control and Prevention and by the Occupational Safety and Health Administration when handling and disposing of infectious agents.

This kit contains materials with small quantities of sodium azide. Sodium azide reacts with lead and copper plumbing to form explosive metal azides. Upon disposal, flush drains with a large volume of water to prevent azide accumulation. Avoid ingestion and contact with eyes, skin and mucous membranes. In case of contact, rinse affected area with plenty of water. Observe all federal, state and local regulations for disposal.

PROCEDURAL NOTES / LAB QUALITY CONTROL

1. Before mixing plexes, **check that each protein is represented by a unique bead region.** Up to 10 bead concentrates (singleplexes or pre-mixed multiplexes) can be combined to increase the number of proteins measured.
2. The **FIDIS™ Human Extracellular Antibody Bead Kit** assay may be combined with other Extracellular Antibody Bead Kits for higher level multiplexing. See the **INFORMATION SHEET and APPENDIX I** for further information. To ensure that several Antibody Bead Kits are compatible in a multiplexed assay, verify that the bead region for each analyte (stated on the **INFORMATION SHEETS** in the Antibody Bead Kits used) is unique.
3. Do not freeze any component of this kit. When not in use, kit components should be stored at +2°C to +8°C. All reagents should be brought to room temperature before use. (air-warm all reagents at room temperature for at least 30 minutes, or alternatively, in a room-temperature water bath for 20 minutes (except plate and standard vials)).
4. **The fluorescent beads are light-sensitive.** Protect the beads from light to avoid photobleaching of the embedded dye. Aluminum foil should be used to cover test tubes used in the assay. Filter plates containing beads should be shielded with an aluminium foil-wrapped plate cover. The

amber vial does not provide full protection, and should therefore be kept covered in the box when not in use.

5. Do not expose beads to organic solvents.
6. Do not place filter plates on absorbent paper towels during loading or incubations, as liquid will be lost due to wicking. An extra plate cover serves as a good surface upon which to rest the filter plate. Following plate washing, excess liquid may be blotted from the bottom of the plate by pressing the plate on clean paper towels.
7. Do not invert the filter plates during the assay. The filter plates are designed to be used in conjunction with a vacuum manifold (DO NOT EXCEED 5 inches Hg) and emptied from the bottom.
8. When pipetting reagents, maintain a consistent order of addition from well-to-well. This method ensures equal incubation times for all wells.
9. Avoid touching the filter plate membrane with pipette tips to prevent tearing.
10. Do not use reagents after kit expiration date.
11. In-house controls should be run with every assay. If control values fall outside pre-established ranges, the accuracy of the assay is suspect.
12. Do not mix or substitute reagents with those from other lots or sources.

PREPARATION OF WASH SOLUTION

Upon storage at +2°C to +8°C, a precipitate may form in the 20x *Wash Solution Concentrate*. If this occurs, warm to +37°C and vortex until the precipitate is dissolved.

The *Wash Solution Concentrate* is provided as a 20x concentrate. To prepare the Working Wash Solution for use with a 96 well plate, transfer the entire contents of bottle to a 500 mL container and add 285 mL of deionized water. If not using an entire 96 well plate, smaller volumes of 1x Working Wash Solution can be made by mixing 1 part of 20x concentrate with 19 parts deionized water.

The Working Wash Solution is stable for up to two weeks when stored at +2°C to +8°C.

Note: To prepare smaller volumes of 1X Working Wash Solution, mix 1 part of 20X concentrate with 19 parts of deionized water. Mix well.

DIRECTIONS FOR WASHING

Incomplete washing will adversely affect assay outcome. All washing must be performed with the Wash Solution provided. All phases of the assay, including incubation steps, washing steps, and loading the beads into the FIDIS™ instrument, are performed in the filter bottom plate provided. Unused wells may be left dry during the assay and used at a later time.

1. To wash beads, place the filter plate on the vacuum manifold and aspirate the liquid with gentle vacuum (**do not exceed 5 mm Hg**). Excessive vacuum can cause the membrane to tear, resulting in antibody bead loss. Prevent any vacuum surge by opening and adjusting the vacuum on the manifold before placing the plate on the manifold surface.
2. Stop the vacuum pressure as soon as the wells are empty. Do not attempt to pull the plate off the vacuum manifold while the vacuum is still on or filter plate damage may occur. Release the vacuum prior to removing the plate.
3. If solution remains in the wells during vacuum aspiration, **do not detach the bottom of the 96 well filter plate.** In some cases, minor clogs in the filter plate may be dislodged by carefully pressing the bottom of the plate under the

clogged well with the pointed end of a 15 mL plastic conical tube. Place the filter plate on a clean paper towel and use a gloved thumb or a 1 mL Pasteur pipette bulb to plunge the top of the clogged well. Empty all clogged wells entirely before continuing the washes. **Note:** Do not attempt to repetitively pull vacuum on plates with clogged wells. This can compromise the unclogged wells and bead loss may occur.

- After all wells are empty, lightly tap or press the filter plate onto clean paper towels (hold the plate in the center for tapping) to remove excess fluid from the bottom of the filter plate. **Do not invert plate.**
- Following the last aspiration and plate taps, use a clean absorbent towel to blot the bottom of the filter plate before addition of next liquid phase or data acquisition step.
- Do not leave plate on absorbent surface when adding reagents.**

PREPARATION OF SAMPLE

- Serum, plasma, and tissue culture medium samples have been evaluated with [bmd](#)'s Multiplex Bead Assays. When possible, avoid the use of hemolyzed or lipemic sera. Suitable sample types are defined on the **INFORMATION SHEET** included in this kit.
- Samples should be collected in pyrogen/endotoxin-free polypropylene tubes. Centrifuge, separate, and transfer samples to polypropylene tubes for storage.
- Samples should be analyzed shortly after collection or frozen. Avoid multiple freeze-thaw cycles of frozen samples. Thaw completely and mix well (DO NOT VORTEX) prior to analysis.
- All samples need to be clarified by centrifugation (1,000 x g for 10 minutes) and/or filtered prior to analysis to prevent clogging of the filter plates.
- Samples with concentrations that exceed the standard curve should be diluted and reanalyzed. Serum or plasma samples should be diluted in *Assay Diluent*. Tissue culture supernatants should be diluted in the corresponding tissue culture medium.

REAGENT PREPARATION

The **FIDIS™ Human Extracellular Antibody Bead Kit** may also be used in conjunction with other bead reagents allowing for a higher level of multiplexing (please see **APPENDIX I** for preparation). Prior to starting the assay, prepare standard and antibody conjugated beads in accordance with instructions below, then proceed to **Assay Procedure, Step 1**.

Preparation of Standard

Each kit comes with 2 complete sets of standard vials, so that 2 runs on the plate can be made with freshly prepared standards.

Reconstitute the protein standard within one hour of performing the assay. All standards are calibrated to NIBSC preparations, when available. Additional standards are available from [bmd](#). The standard included in this kit is provided as a premixed set of related markers.

Before performing standard mixing and serial dilutions confirm reconstitution volumes on the **INFORMATION SHEET** included in the Singleplex Bead Kit(s).

The concentrations of the protein components of the standard are indicated on the **INFORMATION SHEET** included in this kit. Standard dilutions may be performed in glass or polypropylene tubes.

When using serum or plasma samples, reconstitute the standard with *Assay Diluent* provided.

If using other sample types (e.g., tissue culture supernatant), reconstitute the standard with a mixture composed of 50% *Assay Diluent* and 50% of the matrix which most closely matches the sample type (50%/50% mixture).

For example: When the sample type is RPMI medium containing 5% FBS, the standards should be reconstituted in a mixture composed of 50% *Assay Diluent* and 50% RPMI containing 5% FBS.

Reconstitution of Lyophilized Standards

To the standard vial(s) add the suggested reconstitution volume listed below of the appropriate diluent (see above). Do not vortex. When mixing or reconstituting protein solutions, always avoid foaming.

Replace the vial stopper and allow the vial to stand undisturbed for 10 minutes.

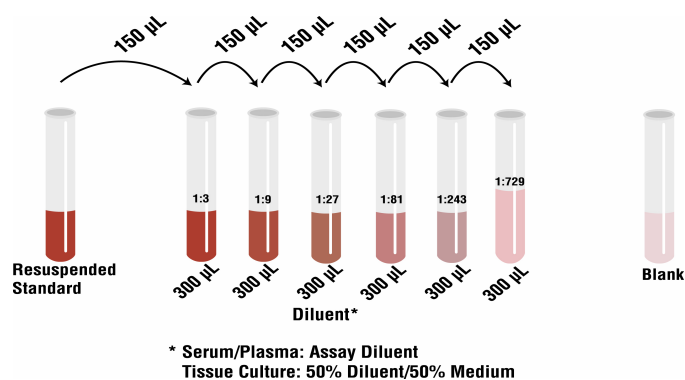
Gently swirl and invert the vial 2-3 times to ensure complete reconstitution and allow the vial to sit for an additional 5 minutes.

Protein standards may be analyzed alone, or may be combined with other protein standards for higher levels of multiplexing. **DO NOT COMBINE MORE THAN 4 VIALS.**

| | |
|--------------------------|---|
| One vial of standard | Reconstitute the standard vial in the suggested reconstitution volume, usually 1 mL, of appropriate diluent (see above). |
| Two vials of standards | Reconstitute each vial with 0.5 mL of appropriate diluent (see above). Combine 300 µL from each vial and mix by pipetting up and down 5-10 times. |
| Three vials of standards | Reconstitute each vial with 0.333 mL of appropriate diluent (see above). Combine 200 µL from each vial and mix by pipetting up and down 5-10 times. |
| Four vials of standards | Reconstitute each vial with 0.250 mL of appropriate diluent (see above). Combine 150 µL from each vial and mix by pipetting up and down 5-10 times. |

Prepare the Standard Curve

The standard curve is made by serially diluting the reconstituted standard in *Assay Diluent* (serum and plasma samples) or a mixture of 50% *Assay Diluent* plus 50% culture medium. See diagram below. **DO NOT VORTEX**. Mix by pipetting up and down 5-10 times.



Discard all remaining reconstituted and diluted standards after completing assay. Return the *Assay Diluent* to the kit.

Preparation of Antibody Conjugated Beads

Determine the number of wells required for the assay. Standard curves and samples may be run singly or in replicates, as desired.

The beads are provided as a 10x concentrate and must be diluted prior to use. The fluorescent beads are light-sensitive. **Protect antibody conjugated beads from light during handling.**

Immediately before dispensing, vortex the 10x bead concentrate for 30 seconds followed by sonication in a sonicating water bath for 30 seconds. To make a 1x stock, dilute 2.5µL from each 10x bead vial in 22.5µL Working Wash Solution per assay well. Each well requires 25µL of the diluted beads. See table below for examples of volumes to combine.

Note: Dilution factor is 1:11 for extra pipetting volume.

| Number of Wells | Vol. 10x Capture Beads (Inflammatory 5-Plex) | Vol. Working Wash Solution |
|-----------------|--|----------------------------|
| 24 | 0.06 mL | 0.6 mL |
| 32 | 0.08 mL | 0.8 mL |
| 40 | 0.10 mL | 1.0 mL |
| 48 | 0.12 mL | 1.2 mL |
| 56 | 0.14 mL | 1.4 mL |
| 64 | 0.16 mL | 1.6 mL |
| 72 | 0.18 mL | 1.8 mL |
| 80 | 0.20 mL | 2.0 mL |
| 88 | 0.22 mL | 2.2 mL |
| 96 | 0.24 mL | 2.4 mL |

ASSAY PROCEDURE

Recommendation : Reverse pipetting

To reduce bubbles and loss of reagents due to residual fluid left in pipette tips, use the recommended reverse pipetting technique.

a. To reverse pipette, set the pipette to the appropriate volume needed.

Note: Do not reverse pipette volumes <20 µL.

b. Press the push-button slowly to the first stop and then press on past it. Note: the amount past the first stop will depend on the volume of liquid available to aspirate from.

c. Immerse the tip into the liquid, just below the meniscus.

d. Release the push-button slowly and smoothly to the top resting position to aspirate the set volume of liquid.

e. Place the end of the tip against the inside wall of the recipient vessel at an angle.

f. Press the push button slowly and smoothly to the first stop. Some liquid will remain in the tip, this should not be dispensed.

g. Remove the tip, keeping the pipette pressed to the first stop.

NOTE: Bring all reagents and samples to room temperature before use.

- Use an adhesive plate cover to seal any unused wells. This will keep the wells dry for future use.
- Pre-wet the wells designated for the assay. Pipette 0.2 mL of *Working Wash Solution* into designated wells.
- Wait 15 to 30 seconds then aspirate the *Working Wash Solution* from the wells using the vacuum manifold.
- Vortex the diluted bead solution for 30 seconds, then sonicate for at least 30 seconds immediately prior to use in the assay.
- Pipette 25µL of the diluted bead solution into each well. Once the beads are added to the plate, keep the plate protected from light.
- Add 0.2mL *Working Wash Solution* to the wells. Allow the beads to soak for 15 to 30 seconds.
- Aspirate the *Working Wash Solution* from the wells by aspiration with the vacuum manifold. Repeat this washing step.
- Blot the bottom of the filter plate on clean paper towels to remove any residual liquid.
Note: Place the filter plate on a plate cover or non-absorbent surface before all incubations.
- Pipette 50 µL *Incubation Buffer* into each well.
- To the wells designated for the standard curve, pipette 100µL of appropriate standard dilution.
- To the wells designated for the sample, pipette 50µL *Assay Diluent* followed by 50µL sample to each well or 50µL in-house controls, if used.
- Cover filter plate containing beads with an aluminium foil-wrapped plate cover. Incubate the plate for 2 hours at room temperature on an orbital shaker. Shaking should be sufficient to keep beads suspended during the incubation (500-600 rpm). Larger radius shakers will need a lower speed and smaller radius shakers will typically handle higher speeds without splashing.

- Ten to fifteen minutes prior to the end of this incubation, prepare the biotinylated detector antibody in accordance with instructions below, then proceed to **Assay Procedure, Step 14**.

• Preparation of Detector Antibody

The *Biotinylated Detector Antibody* is provided as a 10x concentrate and must be diluted prior to use. To make a 1x stock, dilute 10µL from each *10x Biotinylated Detector Antibody Concentrate* vial in 90µL *Biotin Diluent* per assay well. Each well requires 100µL of the diluted Biotinylated Detector Antibody. See table below for examples of volumes to combine.

Note: Dilution factor is 1:11 for extra pipetting volume.

| Number of Wells | Vol. 10x Biotinylated Detector antibody (Inflammatory 5-Plex) | Vol. Biotin Diluent |
|-----------------|---|---------------------|
| 24 | 0.24 mL | 2.4 mL |
| 32 | 0.32 mL | 3.2 mL |
| 40 | 0.40 mL | 4.0 mL |
| 48 | 0.48 mL | 4.8 mL |
| 56 | 0.56 mL | 5.6 mL |
| 64 | 0.64 mL | 6.4 mL |
| 72 | 0.72 mL | 7.2 mL |
| 80 | 0.80 mL | 8.0 mL |
| 88 | 0.88 mL | 8.8 mL |
| 96 | 0.96 mL | 9.6 mL |

- After the 2 hour capture bead incubation, remove the liquid from the wells by aspiration with the vacuum manifold.
- Add 0.2 mL *Working Wash Solution* to the wells. Allow the beads to soak for 15 to 30 seconds, then aspirate with the vacuum manifold. Repeat this washing step.
- Blot the bottom of the filter plate on clean paper towels to remove residual liquid.
- Add 100µL of prepared *1x Biotinylated Detector Antibody* to each well and incubate the plate for 1 hour at room temperature on an orbital shaker. Shaking should be sufficient to keep the beads suspended during incubation (500-600 rpm).
- Prepare **FIDIS™** instrument during this incubation step. Refer to the Information sheet for all bead regions and standard concentration values.
- Ten to fifteen minutes prior to the end of the detector incubation step, prepare the *Streptavidin-RPE* in accordance with instructions below, then proceed to Assay Procedure, Step 20.

• Preparation of Streptavidin-RPE

The *Streptavidin-RPE* is provided as a 10x concentrate and must be diluted prior to use. **Protect Streptavidin-RPE from light during handling.** Dilute 10µL *10x Streptavidin-RPE* in 100µL *Streptavidin-RPE Diluent* per assay well. Each well requires 100µL of the *diluted Streptavidin-RPE*. See table below for examples of volumes to combine.

| Number of Wells | Vol. 10x Streptavidin-RPE | Vol. Streptavidin-RPE Diluent |
|-----------------|---------------------------|-------------------------------|
| 24 | 0.240 mL | 2.4 mL |
| 32 | 0.320 mL | 3.2 mL |
| 40 | 0.400 mL | 4.0 mL |
| 48 | 0.480 mL | 4.8 mL |
| 56 | 0.560 mL | 5.6 mL |
| 64 | 0.640 mL | 6.4 mL |
| 72 | 0.720 mL | 7.2 mL |
| 80 | 0.800 mL | 8.0 mL |
| 88 | 0.880 mL | 8.8 mL |
| 96 | 0.960 mL | 9.6 mL |

- Remove the liquid from the wells by aspiration with the vacuum manifold.
- Add 0.2mL *Working Wash Solution* to the wells. Allow the beads to soak for 15 to 30 seconds, then aspirate with the vacuum manifold. Repeat this washing step.
- Blot the bottom of the filter plate on clean paper towels to remove residual liquid.
- Add 100µL of prepared *1x Streptavidin-RPE* to each well and incubate the plate for 30 minutes at room temperature on an orbital shaker. Shaking should be sufficient to keep the beads suspended during incubation (500-600 rpm).

24. Remove the liquid from the wells by aspiration with the vacuum manifold.
25. Wash the beads by adding 0.2mL *Working Wash Solution* to the wells; allow the beads to soak for 10 seconds, then aspirate with the vacuum manifold. Repeat this washing step two additional times for a total of 3 washes.
26. Blot the bottom of the filter plate on clean paper towels to remove residual liquid.
27. Add 100µL of *Working Wash Solution* to each well. Shake the plates on an orbital shaker (500-600 rpm) for 2-3 minutes to resuspend the beads.
Note: If the plates cannot be read on the day of the assay, they may be covered and stored in a dark location overnight at +2°C to +8°C for reading the following day without significant loss of fluorescent intensity. Aspirate *Working Wash Solution* from stored plates and add 100µL fresh *Working Wash Solution*. Place the plate on an orbital shaker 2-3 minutes at 500-600 rpm prior to analysis.
28. Uncover the plate; insert the plate into the XY platform of the FIDIS™ instrument, and analyze the samples.
29. Determine the concentration of samples from the standard curve using curve fitting software. It is recommended to use the five parameter algorithm with a weighted function (1/y²) depending on the software package used.

LIMITATIONS OF THE PROCEDURE

Do not extrapolate the standard curve beyond the highest or lowest standard point; the dose-response and data collected in these regions may be non-linear and should be considered inaccurate. Note: in some cases, further dilution of the standard beyond 7 points may be possible to extend the low end of the standard curve.

Dilute samples that are greater than the highest standard with *Assay Diluent* or appropriate matrix diluent; reanalyze these samples and multiply results by the appropriate dilution factor.

Samples are diluted in the assay 1:2 (50µL of sample and 50µL of diluent) relative to the standards. Be sure to account for this dilution factor during sample calculations.

The influence of various drugs, aberrant sera (hemolyzed, hyperlipidemic, jaundiced, etc.) and the use of biological fluids in place of serum/plasma and tissue culture medium samples have not been thoroughly investigated. The rate of degradation of analytes in various matrices may not have been investigated. The immunoassay literature contains frequent references to aberrant signals seen with some sera, attributed to heterophilic antibodies. Though such samples have not been seen to date, the possibility of this occurrence cannot be excluded.

**This kit is for research use only.
Not for human therapeutic or diagnostic use.**

PERFORMANCE CHARACTERISTICS

Please refer to analyte specific **INFORMATION SHEET** for performance claims.

APPENDIX I: MULTIPLEXING ASSAYS

Before mixing plexes it is important to check that each analyte is represented by a unique bead region. Up to 10 bead concentrates (singleplexes or pre-mixed multiplexes) can be combined to increase the number of analytes being monitored.

CAPTURE BEAD PREPARATION

Volume from each vial of 10x Bead Concentrate to combine:

$$\frac{0.025 \text{ mL} \times \text{Assay Wells}}{10} = \text{mL per vial}$$

Volume Working Wash Solution:

$$(0.0275 \text{ mL} \times \text{Assay Wells}) - (\text{mL } 10\text{x Capture Bead per vial} \times \text{vials}) = \text{mL}$$

Final Volume of diluted multiplexed Capture Beads:

$$(\text{mL Working Wash Solution}) + (\text{mL } 10\text{x Capture Bead per vial} \times \text{vials}) = \text{mL}$$

Example Calculation: *Combining 5 vials of 10x Capture Bead Concentrate for 48 assay wells.*

Volume from each vial of each 10x Capture Bead Concentrate to combine:

$$\frac{0.025 \text{ mL} \times 48 \text{ Assay Wells}}{10} = 0.120 \text{ mL per vial}$$

Volume Working Wash Solution:

$$(0.0275 \text{ mL} \times 48 \text{ Assay Wells}) - (0.120 \text{ mL } 10\text{x Capture Beads per vial} \times 5 \text{ vials}) = 0.720 \text{ mL}$$

Final Volume of diluted multiplex Capture Beads:

$$(0.720 \text{ mL Working Wash Solution}) + (0.120 \text{ mL } 10\text{x Capture Beads per vial} \times 5 \text{ vials}) = 1.32 \text{ mL}$$

If desired, premixed beads can be stored at +2°C/+8°C until the expiration date printed on the kit box.

BIOTINYLATED ANTIBODY PREPARATION

Volume from each vial of each 10x Biotinylated Antibody to combine:

$$\frac{0.100 \text{ mL} \times \text{Assay Wells}}{10} = \text{mL per vial}$$

Volume Biotin Diluent:

$$(0.110 \text{ mL} \times \text{Assay Wells}) - (\text{mL } 10\text{x Biotinylated Ab. per vial} \times \text{vials}) = \text{mL}$$

Final Volume diluted Biotinylated Antibody:

$$(\text{mL Biotin Diluent}) + (\text{mL } 10\text{x Biotinylated Ab. per vial} \times 10\text{x vials}) = \text{mL}$$

Example Calculation: *Combining 5 vials of 10x Biotinylated Antibody Concentrate for 48 assay wells.*

Volume of each 10x Biotinylated Antibody to combine:

$$\frac{0.100 \text{ mL} \times 48 \text{ Assay Wells}}{10} = 0.480 \text{ mL per vial}$$

Volume Biotin Diluent:

$$(0.110 \text{ mL} \times 48 \text{ Assay Wells}) - (0.480 \text{ mL } 10\text{x Biotinylated Ab. per vial} \times 5 \text{ vials}) = 2.88 \text{ mL}$$

Final Volume diluted multiplexed Biotinylated Antibody:

$$(2.88 \text{ mL Biotin Diluent}) + (0.480 \text{ mL } 10\text{x Biotinylated Ab. per vial} \times 5 \text{ vials}) = 5.28 \text{ mL}$$

If desired, premixed Biotinylated Detector Antibody can be stored at +2°C - +8°C until the expiration date printed on the kit box.

APPENDIX II: INSTRUMENT SETUP

1. Assign the appropriate **Bead Region** (refer to the kit-specific **Information sheet**) to each analyte.
2. We recommend that the user **count 100 events/bead region**.
3. Set **Minimum Events** to 0.
4. Set **Sample Size** to 50 µl.
5. **Set Flow Rate** to 60 µl/minute.
6. For **FIDIS™** kits we recommend an initial **Double Discriminator (DD)** gate setting of 7,800-15,200. (This setting may vary among instruments and must be determined by the user.)
7. Collect **Median RFU**.

• **Helpful [bmd](#) guides for FIDIS™ Cytokine users**

- Creating a Cytokine method, *April 2006*
- How to run Cytokine, *April 2006*

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PROTOCOL SUMMARY

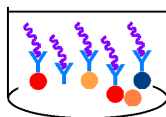
Pre-wet plate
Add 25µL 1x Antibody Coated Beads
200µL Wash Solution

↓ Wash 1 x 200µL

Sample type:

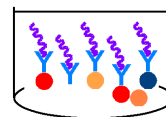
Standard

Serum/Tissue Culture Supernatant



Add 50µL Incubation Buffer
 Add 100µL Standard

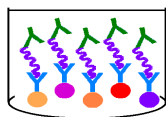
Add 50µL Incubation Buffer
 Add 50µL Assay Diluent



Add 50µL Sample

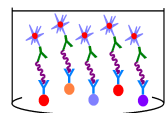


↓ Shake for 2 hours at R.T in the dark
 Wash 2 x 200µL



Add 100µL 1 x Biotinylated Detector Antibody

↓ Shake for 1 hour at R.T in the dark
 Wash 2 x 200µL



Add 100µL 1 x SAV-RPE

↓ Shake for 30 min. at RT in the dark
 Wash 3 x 200µL







Add 100µL Wash Buffer

↓ Shake for 2-3 min

Read in FIDIS™ system

Total time: 3.5 hours

| | | | | | | | |
|---|----------|---|------|---|---------|---|--------------------------------|
|  | Cytokine |  | Bead |  | SAV-RPE |  | Biotinylated Detector Antibody |
|---|----------|---|------|---|---------|---|--------------------------------|

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